

ORIGINAL PAPER

R. M. Levin · A. K. Das

A scientific basis for the therapeutic effects of *Pygeum africanum* and *Serenoa repens*

Received: 18 June 1999 / Accepted: 9 December 1999

Abstract In Europe, phytotherapeutic preparations have been prescribed for the treatment of symptomatic benign prostatic hyperplasia (BPH) for over 20 years [1–4]. In these countries, phytotherapeutic preparations represent approximately 1/3 of total sales of all therapeutic agents sold for the treatment of BPH. In France, and other countries, phytotherapeutic preparations are the most widely used drugs for the treatment of BPH. In Asia, Africa, and India, phytotherapy is considered a first-line treatment for BPH and has been utilized effectively for centuries. In the United States, the multi-million dollar sales of phytotherapeutic preparations for “the health of the prostate and bladder” attests to the widespread utilization of these agents [3, 4].

Two of the most popular phytotherapeutic agents that have undergone both clinical studies to determine their efficacy, and have been the subject of basic science studies to identify the mechanism(s) of action are *Pygeum africanum* (Tadenan), an extract from the bark of the African plum tree, and *Serenoa repens* (Permixon), a lipido-sterol extract of dwarf palm. Tadenan and Permixon are registered therapeutic agents of Debat Pharmaceuticals, and Pierre Fabre Medicament, respectively. Manufacture of both preparations are tightly controlled and subjected to strict quality control for stability of component composition. In regard to phytotherapeutic agents, each individual preparation (even from the same plant source) must be considered individually because of differences in the extraction techniques, preparation of products, composition, and biological activities. Thus, the clinical and biological activities of one preparation cannot be extrapolated to

other preparations of the same plant source. Thus, studies described in this review which utilize the preparations that are manufactured by DEBAT (*Pygeum africanum*) or Pierre Fabre Medicament (*Serenoa repens*) are referred to by their trade names, Tadenan and Permixon, to differentiate them from other nonstandardized preparations of the same plants.

Key words *Serenoa repens* · *Pygeum africanum* · Prostate · BPH · Bladder

There are two aspects to the clinical manifestations of BPH. First is the progressive enlargement of the prostate resulting in compression of the urethra and the initiation of both obstructive and irritative symptoms. These include urgency, frequency, nocturia, hesitancy, reduced flow rate, and incomplete emptying [5–7]. The second is the physical response of the bladder to the enlarged prostate, which includes increased bladder wall thickness, decreased compliance, progressive denervation, collagen infiltration, decreased pressure generation and emptying ability, and unstable bladder contractions during filling [8–12]. Although it has been shown by several studies that there may be no direct relationship between the size of the enlarged prostate and the degree of symptoms [13, 14], most urologists and research scientists in the field would agree that the enlarged prostate is involved in the etiology of the symptoms.

There are several proposed mechanisms involved in BPH pathophysiology including: hormonal stimulation of prostate growth, stroma-epithelial interactions, immune – inflammatory mechanisms, and adrenergic stimulation of cell growth and division. Different agents used in the treatment of BPH have different mechanism of action. In general, current therapies are designed to act at specific aspects of prostate growth and internal prostatic tension, or at the bladder and bladder neck to reduce urethral resistance or reduce the degree of hyperactivity [7, 15, 16]. Finasteride (Proscar) is a 5 α -reductase inhibitor which reduces prostatic dihydrotestosterone (DHT) production. Use of finasteride

R. M. Levin (✉)
Division of Basic and Pharmaceutical Sciences
Albany College of Pharmacy, 106 New Scotland Avenue,
Albany, NY 12208, USA

A. K. Das
Division of Urology Albany Medical College,
Albany, New York, USA

results in a shrinkage of the prostate, theoretically reducing the degree of obstruction present [17, 18]. Several α -adrenergic antagonists are also available for the treatment of symptomatic BPH [7, 15, 16]. There are high densities of α -adrenergic receptors on the smooth muscle components of the prostate, as well as within the smooth muscles of the internal urethral sphincter and urethra. These agents are believed to improve symptomatic BPH by both reducing the internal tension within the prostate and reduce urethral resistance, thus reducing the level of obstruction. Calcium channel blockers and potassium channel openers have been used with limited success to treat the unstable bladder contractions which contribute to the symptoms of BPH [7, 15, 16].

Clinical studies demonstrate that Tadenan and Permixon are statistically more effective than placebo in relieving urological symptoms, and are similar in efficacy in improving urological function to other medications [19–23].

Although still controversial, it is the opinion of the authors, and of many others, that the clinical studies support the clinical effectiveness of these agents in the treatment of symptomatic BPH. Unlike other agents utilized in the treatment of BPH, phytotherapeutic preparations have not been designed to act at any one specific aspect of either prostatic involvement in the etiology of BPH, or the bladder's response to the presence of the enlarged prostate. The end result is that many physicians now accepted that phytotherapeutic agents are clinically effective in the treatment of symptomatic BPH; the mechanism(s) of action are not well understood.

Of the many phytotherapeutic preparations available, there is mechanistic information available only for *Pygeum africanum* and *Serenoa repens*. The following is a review of the pharmacological actions of these phytotherapeutic agents that may be relevant to their clinical efficacy. It should be noted that most of these studies utilize the identical preparations that are currently used as the prescribed therapeutic agents. As mentioned above, bladder dysfunction secondary to BPH directly involves two inter-related processes, the progressive enlargement of the prostate, and the pathological response of the bladder to the enlarged prostate and concomitant

increased urethral resistance. The studies discussed below have been directed at determining the effect of these agents on specific aspects of the growth of the prostate or the response of the bladder to partial outlet obstruction.

***Pygeum africanum* (Tadenan)**

There is evidence that prostatic growth related to BPH involves the proliferative action of one or more growth factors on specific cellular elements within the prostate [24–26]. Tadenan has been shown to be a potent inhibitor of both basal growth of prostatic fibroblasts in culture and the stimulatory effects of a variety of growth factors including basic fibroblast growth factor (bFGF), epidermal growth factor (EGF), and insulin-like growth factor (IGF) [26, 27]. Table 1 lists the IC_{50} doses for inhibition of basal and stimulated proliferation by bFGF, EGF, IGF-I, and nongrowth factor mitogens. Tadenan was effective in inhibiting basal cell growth at concentrations as low as 5 μ g/ml, and growth factor-stimulated proliferation at concentrations as low as 1 μ g/ml [26, 27]. It is interesting to note that the IC_{50} values for Tadenan for inhibition of growth factors and mitogen-stimulated proliferation are in the same order of magnitude (between 4.4 and 14.5 μ g/ml). Thus, the action of Tadenan is likely not via direct inhibition at a specific receptor site, but rather related to inhibition of a common pathway by which these growth factors and mitogens stimulate cell proliferation. Since prostate growth and cellular hyperplasia are controlled by multiple factors, an agent that can inhibit more than one of these factors might be more useful than a specific inhibitor of only one factor.

Inflammatory processes are also believed to be involved in the evolution of prostatic growth and BPH symptomology [28, 29]. *Pygeum africanum* extracts have been shown to cause a dose-dependent inhibition of leukotriene synthesis in human polymorphonuclear cell cultures stimulated by the calcium ionophore A 23187. Inhibition of 5-lipoxygenase was demonstrated at *Pygeum africanum* concentrations as low as 3 μ g/ml [30].

Prostate growth is androgen dependent and the volume and morphology of the prostate is significantly

Table 1 Effect of Tadenan on mitogen-induced proliferation and 5 α -reductase activity in vitro

	Dose range μ g/ml	IC_{50} μ g/ml	Reference
Fibroblast proliferation			27
Basal	1–100	14.4 \pm 4.2	
bFGF-stimulated	1–100	12.6 \pm 1.5	
EGF-stimulated	1–100	4.5 \pm 1.7	
IGF-I-stimulated	1–100	7.8–U-2.7	
Fibroblast proliferation			27
Basal	1–100	12.3 \pm 4	
TPA-stimulated	1–100	12.5 \pm 1.9	
PDBu-stimulated	1–100	8.1 \pm 2.6	
5 α -reductase		63	36
		78	37

affected by alterations in circulatory levels of sex hormones [31–33]. *Pygeum africanum* has been demonstrated to have a “phyto-estrogenic” effect (which can either activate or inhibit estrogen receptors) on the prostate, resulting in a significant effect on the morphology of the glandular epithelium of the prostate, presumably due to a weak anti-estrogen effect [34, 35]. In addition to its “phyto-estrogenic” effect, *Pygeum africanum* has an inhibitory effect on 5 α -reductase, although the potency is relatively low compared to its effect on cellular proliferation [36, 37].

In a rat model, prostate hypertrophy/hyperplasia was stimulated by DHT administration resulting in significant alterations in bladder function. Tadenan pretreatment was shown to significantly improve bladder function simultaneous with reducing prostate weight [38].

In the rabbit, and in man, bladder function can be characterized by the response to autonomic stimulation [39–42]. In the presence of an enlarged prostate (man) and partial outlet obstruction (rabbit), bladder function can remain relatively “normal” for prolonged periods of time during the progression of response to obstruction. This is because the bladder can compensate for the progressive increase in urethral resistance by increasing its mass. During this compensated period of functioning, although there may be changes in micturition pressure and flow characteristics, the urodynamic changes are not severe, and thus do not require medical attention. It is not until decompensated function occurs that the patient seeks medical attention [5–7, 39–41]. These clinical changes may leave the patient susceptible to subsequent renal injury and frequent infections, in addition to the considerable discomfort experienced prior to and during urination.

In man, it is difficult to investigate the cellular mechanisms by which progressive bladder dysfunction occurs. However, many of the functional changes associated with human bladder pathology can be induced in experimental animal model systems and this has been demonstrated prominently in a rabbit model of bladder outlet obstruction [41–43]. The progressive response to partial outlet obstruction can be divided into three distinct phases: (1) an initial response to surgical induction of partial outlet obstruction (days 1–14) characterized by bladder distension followed by a progressive increase in bladder mass and specific contractile and metabolic dysfunctions; (2) compensated bladder function immediately follows the “initial response” and lasts an indefinite and variable length of time; this period is characterized by a relatively stable bladder mass, and stable contractile responses to field stimulation (FS), bethanechol, and KC1 stimulation, but during this period there are progressive morphological changes in bladder cell structure; and (3) at some point, the functional ability to empty degenerates and the bladder becomes “decompensated”. This decompensated phase is characterized by a progressive deterioration in contractility and function (i.e., ability to empty), a rapid

increase in mass, and a progressive decrease in the volume fraction of smooth muscle elements within the bladder wall. The end result is either an organ with a thick fibrous wall, small capacity, poor compliance, and little or no contractile function, or a dilated bladder with a thin fibrous wall, large capacity, and little or no contractile function [41–43].

The following are characteristics of the rabbit model of partial outlet obstruction: The weight of the bladder increases rapidly over the first 2–4 weeks, and then slowly thereafter. This is accompanied by a decreased compliance and a progressive decrease in the contractile responses to all forms of in vitro stimulation [41–43]. However, the rate of contractile dysfunction in response to FS (mediated by the release of neurotransmitters) is faster than the rate of contractile dysfunctions in response to all other forms of stimulation [41–43].

Virtually all bladder cellular elements are involved in the response to partial outlet obstruction [43]. The urothelium undergoes an early rapid period of hyperplasia, the connective tissue component undergoes rapid changes in localization and content, the smooth muscle elements undergo both hyperplasia and hypertrophy, and fibroblasts differentiate into functional smooth muscle cells [44–47]. In general, partial outlet obstruction induces a rapid and marked remodeling of the bladder wall which includes compartment-specific cellular hypertrophy (growth and enlargement of individual cells), hyperplasia (cell division), and re-organization (structural relationship between connective tissue and smooth muscle elements).

We have identified in the rabbit three major cellular alterations which are induced by partial outlet obstruction and directly result in the reduction in the ability of the hypertrophied bladder to generate pressure and empty (1) progressive denervation [48, 49], (2) selective dysfunction of the sarcoplasmic reticulum (SR) to store and release calcium [50, 51], and (3) selective mitochondrial dysfunction [51, 52]. In men with obstructive dysfunctions secondary to BPH, there is a significant increase in bladder mass and wall thickness [54], selective denervation [55, 56], and specific decreases in mitochondrial and SR function [57], indicating that the rabbit model of partial outlet obstruction is a relevant model of obstructive dysfunction secondary to BPH in men.

Several sets of studies on the effects of Tadenan on the response of the rabbit bladder to partial outlet obstruction have been published [58–60]. In the first, rabbits were pretreated for three weeks with vehicle, 1 mg/kg, 10 mg/kg, or 100 mg/kg Tadenan orally [58]. After the pretreatment period, the rabbits were subjected to partial outlet obstruction and continued treatment for an additional two weeks. At the end of 2 weeks’ obstruction, the rabbits were euthanized, the bladders excised and weighed, and in vitro contractility studies were performed. Nonobstructed rabbits were treated with Tadenan or vehicle for 5 weeks [58]. Tadenan had

no effects on bladder mass in obstructed rabbits. In obstructed rabbits, the contractile response of bladder strips to FS and bethanechol was impaired. Tadenan pretreatment resulted in a dose-dependent protection of the contractile responses of bladder strips to FS and bethanechol [58]. Tadenan administration to nonobstructed animals had no effects on any parameter studied.

The second series of experiments determined the effects of Tadenan pretreatment (100 mg/kg) on rabbits partially obstructed for 1–14 days [59]. Similar to the first series of studies, Tadenan had no effects on bladder weights of the obstructed rabbits. Tadenan pretreatment prevented or reduced the severity of the contractile dysfunctions associated with partial outlet obstruction. In addition, Tadenan pretreatment prevented the development of both mitochondrial (citrate synthase) or SR (Ca^{2+} ATPase) dysfunction [59]. In addition, it was shown that Tadenan pretreatment resulted in an overexpression of HSP-70, which is believed to be involved in the protective effect of Tadenan on bladder function [60].

In the next two series of studies [61], Tadenan was administered after 2 weeks of partial outlet obstruction. In the mild obstruction studies, the bladder weight of the Tadenan treated obstructed rabbits was lower than in the vehicle-treated obstructed rabbits. Tadenan treatment reversed the contractile dysfunctions to FS and also reversed the reduction in compliance induced by partial outlet obstruction. In the severe model of partial outlet obstruction, the increase in bladder was greater in the vehicle-treated group than in the Tadenan treated group. Partial outlet obstruction reduced compliance equally in both the vehicle and Tadenan treated groups. Tadenan partially reversed the contractile dysfunction to FS and nearly completely reversed the contractile dysfunction to carbachol [61]. The conclusion from these studies is that Tadenan pretreatment prevented or reduced the progression from compensated bladder function to decompensated bladder function, and Tadenan therapy completely reversed the effects of mild partial outlet obstruction, including improving the compliance of the obstructed bladders; and improved the contractile responses of the severely obstructed bladders. These results are consistent with the theory that Tadenan is acting directly on the bladder by either preventing or delaying the shift from compensated to decompensated function.

Additional studies demonstrated that the reversal of the contractile dysfunctions induced by mild partial outlet obstruction were paralleled by a reversal of the molecular changes to smooth muscle myosin isoforms induced by partial outlet obstruction [62].

Using an established mathematical model of micturition which evaluates the pressures at the bladder neck and the prostatic urethra, Valentini et al. demonstrated that Tadenan administration results in a significant increase in detrusor contractile function that is consistent with the animal data described above [63, 64].

There is evidence that the shift from compensated to decompensated bladder function associated outlet obstruction results from cyclical ischemia and reperfusion [64–66]. Specifically, partial outlet obstruction has been demonstrated to stimulate proteolytic and hydrolytic enzymes [64], which cause progressive damage to specific neuronal and cellular membranes resulting in denervation and mitochondrial and SR damage. The current theory is that Tadenan protects membranes from hydrolytic damage from the activated enzymes [57, 58–60].

If this hypothesis is true and one of the mechanisms of action of Tadenan is protection against ischemic-induced injury, then pretreatment with Tadenan should prevent or reduce the level of injury in an ischemic model of bladder dysfunction. A recently completed study was designed to determine if Tadenan pretreatment of rabbits protected the bladder against the development of contractile dysfunctions induced by unilateral ischemia, and to correlate the contractile effects in the presence and absence of Tadenan with the activation of Hsp-70 [68]. These studies demonstrated that Tadenan pretreatment protected the nonischemic side of the bladder from the development of contractile dysfunctions. In addition, unilateral ischemia (and partial outlet obstruction [60]) resulted in a significantly greater expression of Hsp-70 both in the bladders isolated from the Tadenan treated rabbits and those from the vehicle-treated rabbits [68]. These results indicate that Tadenan has a protective effect against ischemic damage to the bladder, similar to that shown for the obstructed bladder, and this protection may involve an enhanced expression of Hsp-70 and other genetic factors.

Summary

Tadenan has multiple effects on a variety of systems relevant to both prostate growth and bladder function. The summation of these effects would be a reduced growth potential in the prostate and improved bladder function in the presence of mild-moderate obstructive symptoms. The most obvious question concerns whether each of the individual specific actions of Tadenan is mediated by a single component or a select combination of components.

***Serenoa repens* (Permixon)**

Permixon is a lipido-sterolic extract of *Serenoa repens*. Studies have demonstrated that it has three relevant biological activities: antiandrogenic, antiproliferative, and anti-inflammatory. The 5α -reductase is a membrane bound enzyme which catalyzes the reduction of testosterone into its active form dihydrotestosterone (DHT). The prostate is androgen sensitive, and the growth of the prostate in BPH is linked in part to DHT-mediated cell proliferation [31–33]. The source of the DHT is both

from the circulation and from 5 α -reductase activity of the prostate [31–33]. It is clear from the many studies on finasteride, a specific inhibitor of 5 α -reductase type 2, that inhibition of the conversion of testosterone to DHT can significantly reduce the prostate volume in men with symptomatic BPH [69–71].

There are two major isoforms of 5 α -reductase in the human prostate, identified as type 1 and type 2 [72, 73]. The isoforms have different kinetics, pH activity curves, and sensitivities to drugs [72, 73]. Finasteride is a competitive inhibitor of 5 α -reductase type 2, whereas Permixon is a noncompetitive inhibitor of types 1 and 2. Finasteride has a 50-fold greater sensitivity for type 2 5 α -reductase, whereas Permixon has approximately equal potencies for both isoforms. Since Permixon contains multiple components, one cannot directly compare the potencies of finasteride against Permixon. However, the K_i of Permixon for type 1 5 α -reductase is 7.2 μ g/ml and for type 2 is 4.9 μ g/ml [72, 73] (see Table 2). In other studies, extracts of *Serenoa repens* have been shown to inhibit 5 α -reductase activity in extracts from human prostatic epithelium and stroma [74] and significantly interfere with testosterone metabolism [75].

Further studies on the effect of the lipo-sterol extract from Permixon demonstrated that although unsaturated free fatty acids can inhibit type 2 5 α -reductase and saturated free fatty acids can inhibit type 1 5 α -reductase, only the mixture of components of Permixon was able to inhibit both forms [76]. Recent evidence indicates that Permixon may act at the nuclear level to noncompetitively inhibit the enzyme, and may have a selectivity for the prostate over other tissues [77].

The rat prostate is androgen sensitive [78, 79]. Castration results in a substantial shrinkage of the prostate; testosterone administration to the castrate restores prostate volume. In a recent study, Paubert-Braquet demonstrated that the administration of Permixon to castrated rats receiving testosterone significantly inhibited the growth of the prostate, demonstrating that Permixon by its actions on 5 α -reductase inhibited androgen-stimulated growth of the prostate [79].

Recent studies in man demonstrated that three month treatment with Permixon significantly decreased DHT content of the prostate (biopsy analysis) with an asso-

ciated increase in testosterone and decrease in EGF [80]. These changes in testosterone, DHT, and EGF content of biopsies of BPH-prostate tissue confirm the ability of Permixon to inhibit DHT synthesis in vivo. Additional in vivo studies demonstrated that Permixon may stimulate apoptosis and inhibit proliferation [81].

As previously mentioned, the effects of specific growth factors on prostate cells are believed to be involved in the etiology of BPH [24–26]. Permixon at concentrations of 30 μ g/ml, significantly inhibited the ability of bFGF and EGF to stimulate hyperplasia in a primary culture of prostate epithelial cells. However, basal proliferation was not affected [82, 83]. In other in vitro studies, Permixon was shown to be a potent noncompetitive inhibitor of lipoxygenase [84] and cyclooxygenase, both of which are involved in inflammatory processes. As indicated above, infiltration of inflammatory mediators are believed to play a major role in the etiology of BPH through the action of chemotactic mediators such as leukotrienes [28, 29], and recent in vivo studies demonstrate that treatment with Permixon reduces the accumulation of mast cells in the central region of the prostate (rats) [85]. These results are consistent with the idea that Permixon may act in part by reducing the influence of these inflammatory mediators by inhibiting the synthesis of leukotrienes and arachidonic acid metabolites.

In addition to androgen control of prostate growth, prolactin (PRL) has also been demonstrated to significantly stimulate prostate cell hyperplasia [86]. The lipido-sterolic extract from *Serenoa repens* (LSESr) has been shown to be a potent noncompetitive inhibitor of the PRL receptor. Both PRL-stimulated K^+ conductance and Ca^{2+} mobilization in Chinese hamster ovary cells were significantly inhibited by concentrations of LSESr as low as 1 μ g/ml [87].

Distribution studies of ^{14}C -oleic acid or ^{14}C -sitosterol-supplemented *Serenoa repens* in rats demonstrated that both of these components of Permixon are accumulated in the prostate to a greater extent than in the seminal vesicles, brain, or urinary bladder [88]. Since the prostate is a glandular tissue, it would be expected that lipophilic substances would accumulate in the prostate to a greater extent than the bladder, seminal vesicles or

Table 2 Effect of Permixon on hormones, enzyme activities and mitogen-induced proliferation in vitro

	Dose range tested μ g/ml	IC ₅₀ μ g/ml	Competitive / noncompetitive	Reference
5 α -Reductase				
Type 1	0.1–1000	4 \pm 1	Noncompetitive	72, 73
Type 2	0.1–1000	7 \pm 2	Noncompetitive	
Binding of DHT to receptor	1–1000	370	Competitive	80
Lipoxygenase	0.5–50	13		84
Cyclooxygenase (prostate primary culture proliferation)				
Basal	1–30	NS		83
bFGF-stimulated	1–30	App 30		
EGF-stimulated	1–30	App 30		
Prolactin signal transduction	1–30	1	Noncompetitive	87

brain. Although accumulation in a tissue does not necessarily equate to specific activities in this tissue, if the activities of the agent are related to accumulation in specific membranes, and the activity is through localization within the membrane, then this distribution study may be very important.

Summary

Permixon has been shown to be a potent inhibitor of a variety of enzymes and cellular processes which have been implicated in the etiology of BPH. The question of which components of Permixon are responsible for which action is of interest, but does not take away from the specific nature of the actions described above. For example, finasteride is a selective competitive inhibitor of 5 α -reductase, whereas Permixon is a noncompetitive inhibitor.

Finasteride has no specific cellular actions other than its inhibition of 5 α -reductase whereas Permixon has a variety of other actions. The end result is that both agents inhibit the conversion of testosterone to DHT. The advantage of Permixon may be that in addition to inhibiting 5 α -reductase, it also can inhibit growth factor-stimulated proliferation within the prostate and inhibit lipoxigenase activity.

These pharmacological activities of Permixon may in part explain the clinical efficacy of this agent in the treatment of BPH patients.

Summary and conclusions

Tadenan and Permixon have significant effects on a variety of prostate and bladder relevant cellular and biochemical systems. The fact that these agents are noncompetitive inhibitors of several different enzyme systems localized in the prostate indicates that their mechanisms of action are probably not the result of specific receptor or substrate-binding linked mechanisms. It is possible that their ability to inhibit several enzyme systems at similar concentrations is related to the lipophilic nature of the components of these preparations and thus kinetically they appear as noncompetitive inhibitors. The anti-proliferative actions of both Tadenan and Permixon may also be related to the lipophilic nature of the components and their penetration and accumulation within specific cellular and subcellular membranes which results in interference with the ability of a variety (but not all) growth factors to stimulate cell proliferation. The demonstration that many of the actions of both preparations occur at similar concentrations (between 1 and 30 $\mu\text{g/ml}$) is consistent with the above hypothesis.

Tadenan can both prevent and reverse the bladder dysfunctions induced by partial outlet obstruction in rabbits, and these physiological effects may be directly

related to the protection of specific cellular and subcellular membranes from hydrolysis by activated hydrolytic enzymes. One common link between the effects on the prostate and on the bladder is the protection of membranes from the action of specific enzymes. The effect of Permixon on the obstructed bladder is currently under investigation.

Unlike therapeutic agents that are designed to have one mechanism of action, it is likely that phytotherapeutic agents have multiple mechanisms of action. Since there are multiple pathophysiological mechanisms involved in the etiology of symptomatic BPH, the phytotherapeutic agents might be effective in a wider range of patients than conventional therapy.

References

1. Buck AC (1996) Phytotherapy for the prostate. *Br J Urol* 78: 325–336
2. Lowe FC, Dreikom K, Borkowski A, Braeckman J, Denis L, Ferrari P, Gerber G, Levin R, Perrin P, Senge T (1998) Review of recent placebo-controlled trials utilizing phytotherapeutic agents for treatment of BPH. *Prostate* 37: 187–193
3. Fitzpatrick JM, Braeckman J, Denis L, Dreikorn K, Khoury S, Levin R, Perrin P (1996) The Medical Management of BPH with agents other than hormones or alpha-blockers. In: Cockett, Khoury, Chatelain, Denis, Griffiths, Murphy (eds) *Proceedings from the 3rd International Consultation on Benign Prostatic Hyperplasia* (Eds.). Scientific Communications, pp 489–494
4. Dreikorn K, Borkowski J, Braeckman J, Denis L, Gerber G, Levin R, Lowe F, Perrin P, Senge T (1983) Other medical therapies. In: Denis, Griffiths, Khoury, Cockett, McConnell, Chatelain, Murphy, Yoshida(eds) *Proceedings from the 4th International Consultation on Benign Prostatic Hyperplasia*. Scientific Communications pp 635–659
5. Sterling AM, Ritter RC, Zinner NR (The physical basis of obstructive uropathy). In: *Benign prostatic hypertrophy*. Springer, Berlin Heidelberg New York, pp 433–442
6. Grayhack JT, Kozlowski JM (1987) Benign prostatic hyperplasia. In: Gillenwater JY, Grayhack JT, Howards SS, Duckett JW (eds.) *Adult and pediatric urology*. Year Book, Chicago, pp 1062–1126
7. Steers WD, Barrett DM, Wein AJ (1996) Voiding dysfunction: diagnosis, classification and management. In: Gillenwater JY, Grayhack JT, Howards SS, Duckett JW (eds.) *Adult and pediatric urology*, 3rd edn. Year Book, Chicago, pp 1220–1325
8. Levin RM, Haugaard N, O'Connor L, Buttyan R, Das AK, Dixon JS, Gosling JA Obstructive response of human bladder to BPH vs., rabbit bladder response to partial outlet obstruction: a direct comparison. *Neurourol Urodyn* (in press)
9. Kojima M, Inui E, Ochiai A, Naya Y, Ukimura O, Watanabe H (1996) Ultrasonic estimation of bladder weight as a measure of bladder hypertrophy in men with infravesical obstruction: a preliminary report. *Urology* 47: 942–947
10. Manier C, Carte SS, Romano G, Trucchi A, Valent M, Tubaro A (1998) The diagnosis of bladder outlet obstruction in men by ultrasound measurement of bladder wall thickness. *J Urol* 159: 761–765
11. Gilpin SA, Gosling JA, Barnard RJ (1985) Morphological and morphometric studies of the human obstructed, trabeculated urinary bladder. *Br J Urol* 57: 525–529
12. Gosling JA, Dixon JS (1980) Structure of trabeculated detrusor smooth muscle in cases of prostatic hypertrophy. *Urol Int* 35: 351–355
13. Ichiyanagi, O, Nakada T (1997) Correlations between parameters in pressure-flow analysis and histological compositions in

- prostate in patients with benign prostatic hyperplasia. *Urol Int* 59(3): 154–160
14. Terris MK, Afzal N, Kabalin JN (1998) Correlation of transrectal ultrasound measurements of prostate and transition zone size with symptom score, bother score, urinary flow rate, and post-void residual volume. *Urology* 52(3): 462–466
 15. Wein AJ, Longhurst PA, Levin RM (1994) Pharmacologic treatment of voiding dysfunction. In: Mundy AR, Stephenson TP, Wein AJ (eds) *Urodynamics: principles, practice, and application*. Churchill Livingstone, New York, pp 43–70
 16. Holtgrewe HL (1998) Current trends in management of men with lower urinary tract symptoms and benign prostatic hyperplasia. *Urology* 51: 1–7
 17. Olsson GP, Hermamr D, Hammarlund-Udenaes M, Karlsson MO (1999) Validation of a population pharmacokinetic/pharmacodynamic model for 5 α -reductase inhibitors. *Eur J Pharmacol* 8: 291–299
 18. Weisser H, Krieg M (1998) In vitro inhibition of androstenedione 5 α -reduction by finasteride in epithelium and stroma of human benign prostatic hyperplasia. *J Steroid Biochem Mol Biol* 67: 49–55
 19. Barlet A, Albrecht J, Aubert A, Fischer M, Grof F, Grothuesmann HG, Masson J-C, Mazeman E, Mermon R, Reichelt H, Schonmetzler F, Suhler A (1990) Efficacy of *Pygeum africanum* extract in the treatment of micturitional disorders due to benign prostatic hyperplasia: evaluation of objective and subjective parameters: a multicentre, placebo-controlled double-blind trial. *Wien Klin Wochenschr* 232
 20. Breza J, Dzurny O, Borowka A, Hanus T, Petrik R, Blane G, Chadha-Boreham H (1998) Efficacy and acceptability of tadenan (*Pygeum africanum* extract) in the treatment of benign prostatic hyperplasia (BPH): a multicentre trial in central Europe. *Curr Med Res Opin* 14: 127–139
 21. Carraro J C, Raynaud JP, Koch G, Chisholm GD, Di Silverio F, Teillac P, Calais da Silva F, Cauquil J, Chopin DK, Hamdy FC, Hanus M, Hauri D, Kalinteris A, Marencak J, Perier Perrin P (1996) Comparison of phytotherapy (Permixon) with finasteride in the treatment of benign prostatic hyperplasia: a randomized international study of 1098 patients. *Prostate* 29: 231–240
 22. Descotes JL, Rambeaud JJ, Faure G (1996) Placebo-controlled evaluation of the efficacy and tolerability of Permixon in benign prostatic hyperplasia after exclusion of placebo responders. *Clin Drug Invest* 9: 291–297
 23. Grasso M, Montesano A, Buonaguidi A, Castelli M, Lania C, Rigatti P, Rocco F, Cessna BM, Borghi C (1995) Comparative effects of alfuzosin versus Serenoa repens in the treatment of symptomatic benign prostatic hyperplasia. *Arch Esp Urol* 48: 97–103
 24. Iwamura M, Koshiba K, Cockett AT (1998) Receptors for BPH growth factors are located in some neuroendocrine cells. *Prostate Suppl* 8: 14–17
 25. Desgrandchamps F (1997) Clinical relevance of growth factor antagonists in the treatment of benign prostatic hyperplasia. *Eur Urol* 32 [Suppl]: 28–31
 26. Lawson RK (1997) Role of growth factors in benign prostatic hyperplasia. *Eur Urol* 32 [Suppl]: 22–27
 27. Yablonsky F, Nicolas V, Riffaud J-P, Bellamy F (1997) Anti-proliferative effect of *Pygeum africanum* extract on rat prostatic fibroblasts. *J Urol* 157: 2381–2387
 28. Chaudry AA, Wahle KW, McClinton S, Moffat LE (1994) Arachidonic acid metabolism in benign and malignant prostatic tissue in vitro: effects of fatty acids and cyclooxygenase inhibitors. *Int J Cancer* 57: 176–180
 29. Anim JT, Udo C, John B (1998) Characterisation of inflammatory cells in benign prostatic hyperplasia. *Acta Histochem* 100: 439–449
 30. Paubert-Braquet M, Cave A, Hocquemiller R, Delacroix D, DuPont C, Hedef N (1994) Effect of *Pygeum africanum* extract on A23187-stimulated production of lipoxygenase metabolites from human polymorphonuclear cells. *J Lipid Mediat Cell Signal* 9: 285–290
 31. Frick J, Aulitzky W (1991) Physiology of the prostate. *Infection* 19 [Suppl]: S115–118
 32. Griffiths K, Eaton CL, Harper ME, Peeling B, Davies P (1991) Steroid hormones and the pathogenesis of benign prostatic hyperplasia. *Eur Urol* 20 [Suppl]: 68–77
 33. Isaacs JT, Coffey DS (1989) Etiology and disease process of benign prostatic hyperplasia. *Prostate Suppl* 2: 33–50
 34. Mathe G, Orbach-Arbouys S, Bizi E, Court B (1995) The so-called phyto-estrogenic action of *Pygeum africanum* extract. *Biomed Pharmacother* 49: 339–40
 35. Mathe G, Hallard M, Bourut CH, Chenu EA (1995) *Pygeum africanum* extract with so-called phyto-estrogenic action markedly reduces the volume of true and large prostatic hypertrophy. *Biomed Pharmacother* 49: 341–343
 36. Rhodes L, Primka RL, Berman C, Vergult G, Gabriel M, Pierre-Malice M, Gibelin B (1993) Comparison of finasteride (Proscar), a 5 α -reductase inhibitor, and various commercial plant extracts in vitro and in vivo 5 α -reductase inhibition. *Prostate* 22: 43–51
 37. Hartmann RW, Mark M, Soldati F (1996) Inhibition of 5 α -reductase and aromatase by PHL-00801 (Prostatonin), a combination of PY 102 (*Pygeum africanum*) and UR 102 (*Urtica dioica*) extracts. *Phytomedicine* 4: 121–128
 38. Choo M-S, Constantine CE, Bellamy F (1999) Beneficial effects of *Pugeum africanum* extract on dihydrotestosterone induced modifications of micturition and prostate growth in rats (abstract). *J Urol* 161 [Suppl]: 229
 39. Steers WD (1992) Physiology of the urinary bladder. In: Walsh PC, Retik AB, Stamey TA, Vaughan ED Jr (eds) *Cambell's urology*. Saunders, Philadelphia, pp 142–176
 40. Zderic SA, Levin RM, Wein AJ (1996) Voiding function and dysfunction: a relevant anatomy, physiology, and pharmacology, and molecular biology. In: Gillenwater JY, Grayhack JT, Howards SS, Duckett JD (eds) *Adult and pediatric urology* 3rd edn. Year Book, Chicago, pp 1159–1219
 41. Levin RM, Longhurst PA, Monson FC, Kato K, Wein AJ (1990) Effect of bladder outlet obstruction on the morphology, physiology, and pharmacology of the bladder. *Prostate [Suppl]* 3: 9
 42. Levin RM, Monson FC, Longhurst PA, Wein AJ (1994) The rabbit as a model of urinary bladder function. *Neurourol Urodyn* 13: 119–136
 43. Levin RM, Monson FC, Haugaard N, Buttyan R, Hudson A, Roelofs M, Sartore S, Wein AJ (1995) Genetic and cellular characteristics of bladder outlet obstruction. *Urol Clin North Am* 22: 263–283
 44. Levin RM, Monson FC, Haugaard N, Buttyan R, Hudson A, Roelofs M, Sartore S, Wein AJ (1995) Genetic and cellular characteristics of bladder outlet obstruction. *Adv Benign Prostate Hyperplasia* 22: 263–283
 45. Monson FC, McKemra BA, Wein AJ, Levin RM (1992) Effect of outlet obstruction on ³H-thymidine uptake and metabolism: a radiographic and biochemical study. *J Urol* 148: 158–162
 46. Monson FC, Wein AJ, Eika B, Murphy M, Levin RM (1994) Stimulation of the proliferation of rabbit bladder urothelium by partial outlet obstruction and acute overdistension. *Neurourol Urodyn* 13: 51–62
 47. Santarosa R, Colombel M, Kaplan S, Monson F, Levin R, Buttyan R (1994) Hyperplasia and apoptosis: opposing cellular forces that regulate the response of the rabbit bladder to transient outlet obstruction. *Lab Invest*, 70: 503–510
 48. Levitt RM, Saito M, Wein AJ, Packard D, Cohen A, Haugaard M (1993) Effect of partial outlet obstruction on choline acetyltransferase activity in the rat and rabbit. *Neurourol Urodyn* 12: 255–262
 49. Roelofs M, Wein AJ, Barasha B, Monson FC, Passerini-Glazel G, Koteliensky VE, Sartore S, Levin RM (1995) Contractility and phenotype transitions in serosal thickening of obstructed rabbit bladder. *J Applied Physiol* 78: 1432–1441
 50. Zderic S, Rohrmann D, Gong C, Snyder HMcC, Duckett JW, Wein AJ, Levin RM (1996) The decompensated detrusor II: evidence for loss of sarcoplasmic reticulum function following bladder outlet obstruction in the rabbit. *J Urol* 156: 587–592

51. Haugaard N, Wein AJ, Chandy B, Soyupak B, Zderic S, Levin RM (1996) Properties of Ca^{++} - Mg^{++} -ATP-ase in rabbit bladder muscle and mucosa: effect of urinary outlet obstruction. *Neurourol Urodyn* 15: 555-561
52. Haugaard N, Potter L, Wein AJ, Levin RM (1992) Effect of partial obstruction of the rabbit urinary bladder on malate dehydrogenase and citrate synthase activity. *J Urol* 147: 1391-1393
53. Hsu TH-S, Levin RM, Wein AJ, Haugaard N (1994) Alterations of mitochondrial oxidative metabolism in rabbit urinary bladder after partial outlet obstruction. *Mol Cell Biochem* 141: 21-26
54. Kojima M, Inui E, Ochiai A, Naya Y, Ukimura O, Watanabe H (1996) Ultrasonic estimation of bladder weight as a measure of bladder hypertrophy in men with infravesical obstruction: preliminary report. *Urology* 47: 942-947
55. Harrison SCW, Hunnam GR, Farman P, Ferguson DR, Doyle PT (1987) Bladder instability and denervation in patients with bladder outflow obstruction. *Br J Urol* 60: 519-522
56. Gosling JA, Gilpin SA, Dixon JS, Gilpin CJ (1986) Decrease in the autonomic innervation of human detrusor muscle in outflow obstruction. *J Urol* 136: 501-540
57. Levin RM, Haugaard N, Mogavero L, Leggett RE, Das AK (1999) Biochemical evaluation of obstructive bladder dysfunction in men secondary to BPH: a preliminary report. *Urology* 53: 446-450
58. Levin RM, Riffaud J-P, Bellamy D, Rohrmann F, Habib M, Krasnopolsky L, Zhao Y, Wein AJ (1996) Protective effect of Tadenan on bladder function secondary to partial outlet obstruction. *J Urol* 155: 1466-1470
59. Levin RM, Riffaud J-P, Bellamy F, Rohrmann D, Krasnopolsky L, Haugaard N, Zhao Y, Wein AJ (1996) Effects of Tadenan pretreatment on bladder physiology and biochemistry following partial outlet obstruction. *J Urol* 156: 2084-2088
60. Levin ARM, Das AK, Haugaard N, Novitsky Y, Horan P, Leggett RE, Riffaud J-P, Longhurst PA (1997) Beneficial effects of Tadenan therapy following two weeks of partial outlet obstruction in the rabbit. *Neurourol Urodyn* 16: 583-599
61. Chen M-W, Levin RM, Horan P, Buttyan R, (1999) Effect of Tadenan® on the expression of HSP-68 following partial outlet obstruction (abstract). *J Urol* 161 [Suppl]: 229
62. Gomes CM, DiSanto M, Hypolite JA, Horan P, Levin RM, Wein AJ, Chacko S (1999) Reversal of obstruction-induced bladder dysfunction by Tadenan treatment is correlated with a return to the composition of myosin isoform present in the normal detrusor smooth muscle. Proceedings from the 20th Annual Scientific Meeting of the Society for Urodynamics and Female Urology. p P29
63. Valentini FA, Besson GR, Nelson PP (1999) Modelized analysis of the effect of Tadenan (*Pygeum africanum* extract) on the bladder of patients with benign prostatic hyperplasia: Blind vs open study of uroflows. Proceedings from the 20th Annual Scientific Meeting of the Society for Urodynamics and Female Urology. p P2
64. Greenland JE, Hvistendahl JJ, Andersen H, Jorgensen TM, Constantinou CE, McMurray G (1997) Detrusor and kidney blood flow is reduced in response to early bladder outlet obstruction in pigs. *J Urol* 157: 172
65. Azadzi KM, Pontari M, Vlachiotis J, Siroky MB (1996) Canine bladder blood flow and oxygenation: changes induced by tilling, contraction and outlet obstruction. *J Urol* 155: 1459-1465
66. Lin AT, Chen MT, Yang CH, Chang LS (1995) Blood flow of the urinary bladder: effects of outlet obstruction and correlation with bioenergetic metabolism. *Neurourol Urodyn* 14: 285-292
67. Zhao Y, Levin SS, Wein AJ, Levin RM (1997) Correlation of ischemia/reperfusion and partial outlet obstruction induced spectrin proteolysis by calpain with contractile dysfunction in the rabbit bladder. *Urology* 49: 293-300
68. Chen M-W, Levin RM, Horan P, Buttyan R (1999) Effects of unilateral ischemia on the contractile response of the bladder: protective effect of Tadenan. *Mol Urol* 3:5-10
69. Panne J, Marks LS, Pearson JD, Rittenhouse HG, Chart DW, Shety ED, Gormley GJ, Subong EN, Kelley CA, Stoner E, Partin AW (1998) Influence of finasteride on free and total serum prostate specific antigen levels in men with benign prostatic hyperplasia. *J Urol* 159: 449-453
70. Marberger M (1998) Long-term effects of finasteride in patients with benign prostatic hyperplasia: a double-blind, placebo-controlled, multicenter study. PROWESS Study Group. *Urology* 51: 677-686
71. McConnell JD, Bruskewitz R, Walsh P, Andriole G, Lieber M, Holtgrewe HL, Albertsen P, Roehrbom CG, Nickel JC, Wang DZ, Taylor AM, Waldstreicher J (1998) The effect of finasteride on the risk of acute urinary retention and the need for surgical treatment among men with benign prostatic hyperplasia. Finasteride long-term efficacy and safety study group. *N Engl J Med* 338: 557-563
72. Bayne CW, Grant ES, Chapman K, Habib FK Characterisation of a new co-culture model for BPH which expresses 5 alpha-reductase types I and II: The effects of Permixon on DHT formation
73. Iele C, Delos S, Guirou O, Tate R, Raynaud J-P, Martin P-M (1995) Human prostatic steroid 5-reductase isoforms. A comparative study of selective inhibitors. *J Steroid Biochem Mol Biol* 54: 273
74. Weisser H, Tunn S, Behnke B, Krieg M (1996) Effects of the sabal serrulata extract IDS 89 and its subfractions on 5 alpha-reductase activity in human benign prostatic hyperplasia. *Prostate* 28: 300-306
75. Delos S, Carsol JL, Ghazarossian E, Raynaud JP, Martin PM (1995) Testosterone metabolism in primary cultures of human prostate epithelial cells and fibroblasts. *J Steroid Biochem Mol Biol* 55: 375-383
76. Raynaud J-P, Cousse H, Martin P-M (1998) Selectivity of free fatty acids on 5α-reductase inhibitory activity. *J Urol* 159 [Suppl]: 110
77. Bayne CW, Donnelly F, Ross M, Miller WR, Habib F (1999) Differential effects of the BPH drug Permixon on cells from various tissues (abstract). *J Urol* 161 [Suppl]: 228
78. Wright AS, Thomas LN, Douglas RC, Lazier CB, Rittmaster RS (1996) Relative potency of testosterone and dihydrotestosterone in preventing atrophy and apoptosis in the prostate of the castrated rat. *J Clin Invest* 98: 2558-2563
79. Paubert-Braquet M, Richardson FO, Servent-Saez N, Gordon WC, Monge M-C, Bazan NG, Authie D, Braquet P (1996) Effect of *Serenoa repens* extract (Permixon) on estradiol/testosterone-induced experimental prostate enlargement in the rat. *Pharmacol Res* 34: 171-179
80. Di Silverio F, Monti S, Sciarra A, Varasano PA, Martini C, Lanzara S, D'Eramo G, Di Nicola S, Toscano V (1998) Effects of long-term treatment with *Serenoa repens* (Permixon) on the concentrations and regional distribution of androgens and epidermal growth factor in benign prostatic hyperplasia. *Prostate* 37: 77-83
81. Vacherot F, Azzouz M, Gil-Diez-de Medina S, Raynaud JP, Abou CC, Chopin D (1999) Effect of Permixon on apoptosis and proliferation in the prostate of patients with BPH (abstract). *J Urol* 161 [Suppl]: 228
82. Paubert-Braquet M, Raynaud J-P, Braquet PG, Castres HC (1997) Permixon [lipid sterolic extract of serenoa repens (LSEsr)] and some of its components inhibits b-FGF- and EGF-induced proliferation of human prostate organotypic cell lines. *J Urol* 157 [Suppl]: 138
83. Paubert-Braquet M, Cousse H, Raynaud JP, Mencia-Huerta JM, Braquet P (1998) Effect of the lipidosterolic extract of *Serenoa repens* (Permixon) and its major components on basic fibroblast growth factor-induced proliferation of cultures of human prostate biopsies. *Eur Urol* 33: 340-347
84. Paubert-Braquet M, Mencia, Huerta JM, Cousse H, Braquet P (1997) Effect of the lipidic lipidosterolic extract of *Serenoa repens* (Permixon) on the ionophore A23187-stimulated production of leukotriene B4 (LTB4) from human polymor-

- phonuclear neutrophils. Prostaglandins Leukot Essent Fatty Acids 57: 299–304
85. Mitropoulos D, Kiroudi A, Mitsogiannis I, Costomitsopoulos N, Kittas C, Karayannacos P, Dimopoulos C (1999) In vivo effect of the lipido-sterolic extract of *Serenoa repens* (Permixon) on mast cells accumulation and glandular epithelium trophism in the rat prostate (abstract). J Urol 161 [Suppl]: 362
 86. Thomas JA, Keenan EJ (1976) Prolactin influences upon androgen action in male accessory sex organs. Adv Sex Hormone Res 2: 425–470
 87. Vacher P, Prevarskaya N, Skryma R, Audy MC, Vacher AM, Odessa MF, Dufy B (1995) The lipidosterolic extract from *Serenoa repens* interferes with prolactin receptor signal transduction. J Biomed Sci 2: 357–365
 88. Chevalier G, Benard P, Cousse H, Bengone T (1997) Distribution study of radioactivity in rats after oral administration of the lipidosterolic extract of *Serenoa repens* (Permixon) supplemented with [^{14}C]-oleic acid or [^{14}C]-qsitosterol. Eur J Drug Metab Pharmacokinet 22: 1–10

ANNOUNCEMENTS

2000

28th Munich Endourological Symposium with Nursing Staff Seminar 5–6 October 2000, Munich, Germany

Chairman: Prof. Dr. R. Hartung,

Information: Dr. R. Paul, Department of Urology, Technische Universität München, Klinikum rechts der Isar, Ismaninger Str. 22, 81675 München; Tel: 089/41402507, Fax: 089/41402585, e-mail: mriu@lrz.tu-muenchen.de, Internet: <http://www.med.tu-muenchen.de/uro/endo.html>

4th International Symposium on Uro-Onkology: Advances in Diagnosis and Treatment of Prostate Cancer 19–21 October 2000, Marburg, Germany

Topics: Live Surgery (ascending, descending, nerve-sparing retropubic prostatectomy, perineal prostatectomy, laparoscopic prostatectomy, art. Sphincter implantation, penile prosthesis implantation), pathology and molecular biology of PCA, therapeutic options in organ confined PCA, treatment of locally advanced and metastatic PCA.

Information: Priv. Doz. Dr. A. Heidenreich, Department of Urology, Philipps-Universität Marburg, Baldingerstrasse, 35043 Marburg, Germany; Tel: +49 6421 286 6239, Fax: +49 6421 286 5590, e-mail: heidenre@post.med.uni-marburg.de